Thursday, July 03, 2025
Zoom

Meeting Minutes

VOTING MEMBERS PRESENT:	K. Burns, J. Corcoran, M. Espinola, G. Dean, S. Kasper, R. Larson, E. Otten, T. Rausch, F. Schaefer, E. Serafin, J. Yu
VOTING MEMBERS NOT PRESENT:	S. Apewokin, G. Babcock, D. Elsaesser,
AD HOC MEMBERS/CONSULTANTS/GUESTS:	A. Perry, J. Strasser, C. O'Rourke
IBC STAFF:	D. Healy, B. Kesavalu

K. Burns convened the meeting at 12:00 p.m.

I. **Conflicts of Interest** – No conflicts
II. **Minutes** - Minutes from the previous IBC meeting (05/01/25) were approved (11:YES/0:NO/0:Abstained)
III. **Old Business** - No old business was discussed
IV. **New Business**

A. **Primary Protocols** (2 protocols)

IBC Protocol Number	PI's Link	Type of Submission	IBC Items
1. 25-06-04-01	**Steele**	New BSL2	***in vitro:*** HDM (blood) ***in vitro*** and ***in vivo:*** lentiviral and adenoviral vectors, Zika virus, and HDM (PBMCs, cell lines)
IBC Requests:	**IBC Form** 1. **Section I. A** and **I. B** - Provide the Internal Medicine division that you belong; 2. **Section I. A** and **I. B** - Provide UC mail code; 3. **Section I. D** – Edit "CCCMC" to "CCHMC"; 4. **Section II. B** - Indicate the name of the oncolytic virus used and provide the reason it was chosen; 5. **Section II. B -** "human donor patent" should read "human donor patient"; 6. **Section II. B** - **Last sentence** - If lab has written SOPs for different activities, they should be attached to the protocol. If the statement about refers to the safety procedures outlined in this protocol, it needs to be reworded for clarity; 7. **Section III. A - Pipet** - Include aerosol mitigation while pipetting. Information can be found in the eManual on aerosols; 8. **Section III. B -** Check YES for "Additional Information" and indicate which activities involving biological hazardous materials are conducted inside of a fume hood; 9. **Section III. B -** In "Additional Information", list types of sharps that are disposed of in the sharps container; 10. **Section III. B -** In "Additional Information", indicate why safety goggles are used; **Form A – Recombinant or Synthetic Nucleic Acid:** 11. **Section I - Genes #1 to #7 -** In "Additional Information", indicate what tool(s) is used for gene inhibition (e.g. CRISPR/Cas9);		

	12. **Section I - Gene #7 -** In "Additional Information", explain the use of this toolkit; 13. **Section I - Gene #8** - In "Additional Information", indicate the genes that are targets for Cas9; 14. **Section I - Gene #9 -** The first gene listed should be "GFP" Also, confirm the name of the last (3rd) gene; 15. **Section I - Gene #9 - Gene Source** - The source for the second gene is "arthropod". Change selection from "other" to "arthropod"; 16. **Section II. B - Systems #1 and #2** - List the name of the genes that are inhibited. For the neural toolkit just say "neural identity genes"; 17. **Section II. B - System #2 -** "Parent Virus" should have the serotype of the virus that originated the vector (most likely Ad-5); 18. **Section II. B - System #2** – "Expression Construct" is the name of plasmid which contains the adenovirus backbone. Correct entry; **Form B - Microbial/infectious agents** 19. **Agent's Characteristics** - Provide the characteristics of the different ZIKV strains (e.g. pathogenicity, clinical manifestations, type of attenuation); 20. **Use *in vivo*** - Limit information to the administration routes of Zika Virus. Details about animal procedures should be transferred to Form D - Section II - Additional Information (under the table); **Form C – Human and NHP Derived Materials**: 21. **Section I - Table - Cell lines** - Information about mouse cell lines are not relevant to this form. Provide examples of human cells used and from where they were obtained. Also, include "Vero cells" (cell of non-human primate origin) and their source; 22. **Section I - Table** - **Cell lines** - The animal form (Form D) indicates that transduced cells are implanted into animals. If cells are of human origin, "*in vivo*" needs to be selected and Section II. B must be completed; 23. **Section I - Table** - **Blood** - Indicate if PBMCs are obtained from pancreatic cancer patients; 24. **Section II. A -** Include experiments involving human PBMC; **Form D – Biohazard in Animals:** 25. **Section I** - Provide the IACUC protocol number; 26. **Section II - Table -** Select "Virus" instead of "Viral Vector"; 27. **Section II - Animal Species** - "Murine" is too broad. If mice and rats are used, provide the species; 28. **Section III. A -** Include the risks of exposure for animal procedures involving Zika virus; 29. **Section III. C** - In "Additional Information", list the types of sharps that are disposed of in the sharps container.			
Motion	Approve upon modifications addressing requests.			
Voting Result & **Dual Use**	**YES:** 11	**NO**: 0	**Abstained**: 0	**Dual Use**? No

<table>
<tr><th>IBC Protocol Number</th><th>PI's Link</th><th>Type of Submission</th><th>Biosafety Level</th><th>IBC Items</th></tr>
<tr><td>2. 24-01-02-01</td><td>Waltz</td><td>Amendment</td><td>BSL2</td><td><u>in vitro</u>: lentiviral vector, retroviral vector and HDM (established human cell lines)
<u>in vivo:</u> virally transduced cells
AMENDMENT: <u>in vivo</u> adenoviral vector</td></tr>
<tr><td>IBC Requests:</td><td colspan="4">1. Lab staff need to complete their viral vector training;
<u>Main Form – General Safety</u>:
2. Section II. B - 3rd paragraph – Clarify if adenoviral vectors are also used for in vitro work;
3. Section III. A - Sonicator - Replace biosafety "hood" with biosafety "cabinet";
4. Section III. B - Review this section and select any other engineering controls used in the lab while handling biological hazardous materials. If the lab has sharps containers, indicate that they are available in "Additional Information" and indicate the type of sharps used;

<u>Form A – Recombinant or Synthetic Nucleic Acid:</u>
5. Section I- #1 - In "Additional Information", include the other tool that is used to inhibit Ron;
6. Section I- #2 and #3 - Check YES for "Additional Information" and indicate how genes are inhibited (e.g. shRNA, siRNA, miRNA, CRISPR/Cas9);
7. Section I- #4 - "Gene Source" for Cre recombinase is "virus". Correct the field;
8. Section I - #4 – For completion, in "Additional Information", include the gene that Cre targetss (i.e. Ron);
9. Section II. B - System #4 - "Parent Virus" for the system is likely to be "Ad5". Update for correctness;
10. Section II. B - System #4 - "Expression Construct" is the name of the plasmid with the adenovirus backbone. What was provided is the type of deletions of the vector, transfer the information to "Additional Information";
11. Section II. B - System #4 – Select "in vivo" and confirm whether "in vitro" work will occur;
12. Section II. B - System #4 - Gene Inhibited - Considering that Cre expression will cause inhibition of Ron, replace "N/A" with "Ron".

<u>Form B - Microbial/infectious agents</u>
13. Agent #1 - Source - Indicate the correct source instead of "commercially".

<u>Form D – Biohazard in Animals:</u>
14. Section I - Remove the old IACUC protocol number;
15. Section II - Table - Viral Vectors - Animals injected with viral vectors don't need to stay within the ABSL2 for the entire study. Uncheck box in this regard;
16. Section II - Additional Information - Provide the administration route(s) for adenoviral vectors. Also, in the last sentence replace "trasduction" with "transduction";
17. Section III. A - Revise section to include assessment and procedures for adenoviral vector, including measures on preventing needlesticks;
18. Section III. B - Review section and update it if route for adenoviral vector was not already included;
19. Section III. C - In "Additional Information", list the types of sharps that are disposed of in the sharps container.</td></tr>
<tr><td>Motion</td><td colspan="4">Approve upon modifications addressing requests.</td></tr>
<tr><td>Voting Result & Dual Use</td><td>YES: 11</td><td>NO: 0</td><td>Abstained: 0</td><td>Dual Use? No</td></tr>
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B. **Secondary Protocols** (2 protocols)

IBC Protocol Number	PI's Link	Type of Submission	Biosafety Level	IBC Items
1. 25-06-12-01	**Sherman**	Renewal	BSL2	***in vitro:*** HDM (serum, plasma, PBMC, tissue samples from infected patients: HIV, HAV, HBV, HCV, HDV, HEV, Coronavirus and Zika virus)
IBC Requests:	**Main Form – General Safety**: 1. **Section I. D - Type/Function** – Indicate if any listed locations are used for storage; 2. **Section II. A** - Assuming that viruses are not isolated nor cultured, uncheck the "Microbial/Infectious Agents" box; 3. **Section II. B - 2nd paragraph** - Transfer information to Section IV - Transport Method; 4. **Section III. A** - **Pipet** - In addition to the information provided, additional safety measures need to be followed while performing activities with potential of aerosol generation. For information on aerosol mitigation procedures, consult the eManual to update this section; 5. **Section III. C -** Check "face shield" if you are using or remove this description; 6. **Section VIII -** This section is reserved for materials that are not being used in ongoing (nor in the near-future) research. Revise the section if necessary. **Form C – Human and NHP Derived Materials**: 7. **Section I –** "Organoids" should be unchecked, if no longer applicable; 8. **Section I - Under Table** - Since projects involving iPSCs are no longer conducted, check "NO" for pluripotent cells; 9. **Section II. A -** Remove information that is no longer applicable (e.g. use of iPSCs).			
Motion	Approve upon modifications addressing requests.			
Voting Result & **Dual Use**	**YES:** 11	**NO**: 0	**Abstained**: 0	**Dual Use**? No

IBC Protocol Number	PI's Link	Type of Submission	Biosafety Level	IBC Items
2. 25-06-13-01	**Godar**	Renewal	BSL2	***in vivo* and *in vitro:*** Respiratory Syncytial Virus and HDM (established and primary cells, tissues, bodily fluids)
IBC Requests:	**Form B - Microbial/infectious agents:** 1. **Agents Characteristics** – **Last sentence** – "15% sodium hypochlorite" should read "15% household bleach". **Form C – Human and NHP Derived Materials**: 2. **Section I** – **Table** – **Established Cell Lines** – Check the "NHP" box to reflect use of Vero cells; 3. **Section II. B** - Remove #2 since it is not related to this form (human derived materials) **Form D – Biohazard in Animals:** 4. **Section I** - Update the IACUC protocol number; 5. **Section III. B -** Transfer #2 info to the "Chemical - anesthesia" field; 6. **Section III. B -** Confirm that SC and fat pad injections are done with animals under anesthesia. If for those routes, animals are only mechanically restrained, transfer information to the correct field.			
Motion	Approve upon modifications addressing requests.			
Voting Result & **Dual Use**	**YES:** 11	**NO**: 0	**Abstained**: 0	**Dual Use**? No

V. Protocol Updates (May 29th to June 26th) – 35 protocols

1. IBC# 23-09-14-01 - PI: Schutte - Personnel
2. IBC# 23-08-04-23 - PI: Wang Xuefeng - Personnel
3. IBC# 24-06-18-02 - PI: Byrd - Personnel
4. IBC# 23-04-21-01 - PI: Chella Krishnan - Personnel
5. IBC# 25-02-14-01 - PI: Drosatos - Personnel
6. IBC# 22-11-17-01 - PI: Kotagiri - Personnel
7. IBC# 25-02-14-01 - PI: Drosatos - Personnel
8. IBC# 24-06-18-02 - PI: Byrd - Personnel
9. IBC# 24-03-21-01 - PI: Sertorio - Personnel
10. IBC# 24-01-02-01 - PI: Waltz - Personnel
11. IBC# 24-02-23-01 - PI: Apewokin - Personnel
12. IBC# 25-01-07-02 - PI: Wang YG - Personnel, Location
13. IBC# 25-02-23-01 - PI: Grogan - Personnel
14. IBC# 24-02-01-01 - PI: Blackard - Personnel
15. IBC# 23-07-20-02 - PI: Hite - Location
16. IBC# 23-01-06-01 - PI: Xu - Personnel
17. IBC# 25-04-14-01 - PI: Lynch - Personnel
18. IBC# 23-12-19-01 - PI: Huang W - Location, Personnel
19. IBC# 25-03-20-01 - PI: Ifergan - Location
20. IBC# 24-07-08-01 - PI: Yu - Personnel
21. IBC# 25-03-19-02 - PI: Plas - Personnel
22. IBC# 23-09-28-01 - PI: Zimmermann - Personnel
23. IBC# 24-03-13-01- PI: Shirokawa - Personnel
24. IBC# 25-03-19-01 - PI: Owens - Personnel
25. IBC# 22-08-08-01 - PI: Bhattacharya - Personnel
26. IBC# 24-01-22-01- PI: Maria CK - Personnel, Location
27. IBC# 23-05-17-01 - PI: Davidson Sean - Personnel
28. IBC# 22-10-21-01 - PI: Herman - Personnel
29. IBC# 25-03-19-01 - PI: Owens - Personnel
30. IBC# 24-01-09-01 - PI: Lander- Personnel
31. IBC# 24-12-09-01 - PI: Deepe - Personnel
32. IBC# 24-06-18-02 - PI: Byrd - Personnel
33. IBC# 25-02-14-01 - PI: Drosatos - Personnel
34. IBC# 24-01-17-01 - PI: Askew - Personnel and BSL2 upgrade
35. IBC# 22-09-23-02 - PI: Park -Personnel, Secondary contact

VI. **Reports**

A. IBC - FY25 IBC Overview

M. Espinola presented an overview of active IBC protocols highlighting the updates and renewals that occurred during the fiscal year.

B. BSOf - FY25 Lab Inspection Overview

B. Kesavalu presented an overview of FY25 inspection highlighting the common deficiencies and improvements.

VII. **Educational Materials/Updates**

A. **Cadaveric Human Growth Hormone–Associated Creutzfeldt-Jakob Disease with Long Latency Period, United States** CDC, June 2025

B. **Cocaine vaccine, a longshot attempt to quell cravings, yields mixed data in small study** Endpoint News, May 2025

K. Burns adjourned the meeting at 12:43 p.m.