

Institutional Biosafety Committee - Regular Meeting

Thursday, January 08, 2026

Zoom

Meeting Minutes

VOTING MEMBERS PRESENT:	K. Burns, J. Corcoran, G. Dean, D. Elsaesser, M. Espinola, S. Kasper, R. Larson, E. Otten, E. Serafin, T. Rausch, F. Schaefer, J. Yu
VOTING MEMBERS NOT PRESENT:	S. Apewokin, G. Babcock
AD HOC MEMBERS/CONSULTANTS/GUESTS:	T. Gulley, A. Perry, J. Strasser
IBC STAFF:	D. Healy, B. Kesavalu

K. Burns convened the meeting at 12:00 p.m.

- I. **Conflicts of Interest** - No conflicts were identified.
- II. **Minutes** - Minutes from the previous IBC meeting (12/04/25) were approved (10: YES/0: NO/1: Abstained)
- III. **Old Business** - No old business was discussed
- IV. **New Business**
 - A. **Primary Protocols** (0 protocols)
 - B. **Secondary Protocols** (6 protocols)

IBC Protocol Number	PI	Type of Submission	Biosafety Level	IBC Items
1. 25-12-09-01	Dong	Renewal	BSL2	<i>in vitro</i> and <i>in vivo</i> : HDM (cell lines) <i>in vivo</i> : HDM (Patient Derived Xenograft)
IBC Requests:	<p><u>Main Form – General Safety:</u></p> <ol style="list-style-type: none"> Section III. A - Pipet - Change "pitting" to pipetting. In addition to using containment equipment (i.e. BSC), there are safety measures that need to be followed while performing activities with potential for aerosol generation. For information on aerosol mitigation procedures, consult the eManual and update the safety measures; Section III. A - Blender - Include aerosol mitigation strategy for the blender; Section III. A - Sonicator- "solicitor container" - did you mean sonicator? <p><u>Form B – Microbial/Infectious Agents</u></p> <ol style="list-style-type: none"> Agent's Characteristics - Include a brief description of the <i>E. coli</i> species and strains, such as morphology and pathogenicity (and possible antibiotic resistance). <p><u>Form C – Human & Non-Human Primate (NHP) Derived Materials</u></p> <ol style="list-style-type: none"> Section I. A needs to be completed. <p><u>Form D – Biohazard in Animals:</u></p> <ol style="list-style-type: none"> Section II - Additional Information - Form C (section II. B) states that human cells are also transplanted in organs. If that is accurate, describe in this section; Section III. A - Describe what activities offer more risk for exposure (e.g. cell transplantation - needlestick risk); Section III. B - Confirm that anesthesia is used for both organ and subcutaneous inoculation of human cells/pdx. 			
Motion	Approve upon modifications addressing outlined issues.			
Voting Result & Dual Use	YES: 11	NO: 0	Abstained: 0	Dual Use? No

J. Yu joined the meeting at 12:03 p.m.

IBC Protocol Number	PI	Type of Submission	Biosafety Level	IBC Items
2. 25-12-12-01	Wohleb	Renewal	BSL1	<i>in vivo</i> : AAV vector
IBC Requests:	<p><u>Main Form – General Safety:</u></p> <ol style="list-style-type: none"> Section I. C - List phone numbers for all authorized personnel; Section I. D - Section III mentions the use of a cell sorter. Indicate the location of cell sorter in this section; Section III. A - Pipet - Is pipetting used while handling AAV vectors? If not, uncheck box, but if so, describe aerosol mitigation while using pipet aerosols; Section III. A - Cell Sorter - Is a cell sorter used for AAV infected murine cells? If not, uncheck box. <p><u>Form A – Recombinant or Synthetic Nucleic Acid:</u></p> <ol style="list-style-type: none"> Section I - Gene #1 to #5 – “Parent Virus” for AAV vectors is always AAV2. Make this change and in “Additional Information” provide their pseudotype/cap gene (e.g. cap gene of AAV5); Section I - Gene #2 - Include in "Additional Information" that the gene functions as a tumor suppressor; Section I - Gene #5 - Change the spelling of "Phospholipip" to "Phospholipid". <p><u>Form D – Biohazard in Animals:</u></p> <ol style="list-style-type: none"> Section I - Update your IACUC protocol number; Section III. C - It is our understanding that intracranial injections in RC are not performed inside a biosafety cabinet (BSC). If that is accurate and you don't use a BSC, uncheck the box for BSC. 			
Motion	Approve upon modifications addressing outlined issues.			
Voting Result & Dual Use	YES: 12	NO: 0	Abstained: 0	Dual Use? No

IBC Protocol Number	PI	Type of Submission	Biosafety Level	IBC Items
3. 25-12-15-01	McReynolds	Renewal	BSL1	<i>in vivo</i> : AAV vector
IBC Requests:	<p><u>Main Form – General Safety:</u></p> <ol style="list-style-type: none"> Section I. D – The room listed does not have a BSC. Confirm if this LAMS location is used and check NO for BSC . Section III. A - Pipet - In addition to using containment equipment (i.e. BSC), there are safety measures that need to be followed while performing activities with potential for aerosol generation (e.g. use of plugged pipet tips). For information on aerosol mitigation procedures, consult the eManual and update as necessary. <p><u>Form D – Biohazard in Animals:</u></p> <ol style="list-style-type: none"> Section III. C - It is our understanding that in RC intracranial injections are not performed inside a biosafety cabinet (BSC). If that is accurate and you don't use a BSC for other activities in LAMS, uncheck the box for BSC; Section III. C - Because your animal procedures are performed exclusively in LAMS, uncheck the box for "bedding dump stations" since they do not have this type of equipment; Section III. C - Check YES for "Additional Information" and list types of sharps that are disposed of in the sharps containers. 			

Motion	Approve upon modifications addressing outlined issues.			
Voting Result & Dual Use	YES: 12	NO: 0	Abstained: 0	Dual Use? No

IBC Protocol Number	PI	Type of Submission	Biosafety Level	IBC Items
4. 25-12-18-01	Xu	Renewal	BSL2	<u>in vitro</u> : lentiviral vector and HDM (established/packaging cells)
IBC Requests:	<p><u>Main Form – General Safety:</u></p> <ol style="list-style-type: none"> Section I. B - If there is no phone in the lab, enter "N/A"; Section II. A - The only human derived material included in this protocol is HEK 293 cells for production of "pseudovirus" (Form C). Do you also produce lentivirus in your lab? If so, Form A (section II. A) will need to be modified accordingly. If you do not produce viral vectors in your lab nor use HEK cells for other experiments, uncheck the box for human derived materials; Section II. B – Rephrase the paragraph to fix grammatical and spelling errors (e.g. replace "pervious" with "previous") ; Section II. B - For clarity, specify the origin of bone marrow stromal cells (mouse?); Section III. B - Check YES for "Additional Information" and indicate which needle safety and scalpel safety devices are used and what type of sharps will be discarded in Sharps Container. Also, change "Fisher brand sharper containers" to "Fisher brand sharps container". <p><u>Form A – Recombinant or Synthetic Nucleic Acid:</u></p> <ol style="list-style-type: none"> Section II. A - In form C, HEK 293 cells are mentioned as used for making "pseudoviral particles". If that refers to viral vectors and if vector particles are produced in your lab, the "own laboratory" box needs to be selected. <p><u>Form C – Human and NHP Derived Materials:</u></p> <ol style="list-style-type: none"> Section I – If HEK cells are really used, provide their source (ATCC?). 			
Motion	Approve upon modifications addressing outlined issues.			
Voting Result & Dual Use	YES: 12	NO: 0	Abstained: 0	Dual Use? No

IBC Protocol Number	PI	Type of Submission	Biosafety Level	IBC Items
5. 25-12-12-02	Kotagiri	Renewal	BSL2	<u>in vitro</u> : lentiviral vector, HDM (skin)in vitro and <u>in vivo</u> : Influenza virus (H1N1), recombinant <i>Escherichia coli</i> (RG1), <i>Escherichia coli</i> (RG2 - O157:H7), recombinant <i>Staphylococcus epidermidis</i> , <i>Staphylococcus aureus</i> , <i>Klebsiella pneumoniae</i> , <i>Lactobacillus gasseri</i> , <i>Lactobacillus plantarum</i> , <i>Mycobacterium tuberculosis</i> (RG2 - H37Ra), <i>Mycobacterium smegmatis</i> , <i>Pseudomonas aeruginosa</i> , <i>Stenotrophomonas maltophilia</i> , <i>Salmonella typhimurium</i> , <i>Yersinia pseudotuberculosis</i> , mRNA (SARS-Cov2) vaccine, and HDM (established tumor cells)
IBC Requests:	<ol style="list-style-type: none"> Lab staff need to complete their BBP training; <p><u>Main Form – General Safety:</u></p> <ol style="list-style-type: none"> Section I. B - Secondary Contact - Include office and lab phone or enter "N/A"; Section III. B - In "Additional Information" mention that the sonicator will be operated in the fume hood as stated in Section III. A; Section III. C - When are the shoe covers used? Uncheck the "shoe cover" box if that is only used at LAMS; 			

	<p>5. Section V - It is standard practice to disinfect both biosafety cabinets and other surfaces with 10% household bleach followed by 70% alcohol to prevent corrosion. Modify your statement accordingly;</p> <p>6. Section VIII - This section is reserved for materials that are not being used in ongoing (nor in the near-future planned) research. Revise the section if necessary.</p> <p>Form A – Recombinant or Synthetic Nucleic Acid:</p> <p>7. Section I - Form B (microbial agent form) mentions recombinant <i>S. epidermidis</i> expressing Shinorine and Gadusol. If production of this recombinant bacterium is performed in your lab, include these genes in this section;</p> <p>8. Section I - Form D (animal form) mentions about the ischemia & burn models where RG1 bacterium expresses MAM. If production of recombinant bacteria is performed in your lab, include MAM in this section.</p> <p>Form B – Microbial/Infectious Agents</p> <p>9. For all the RG2 agents, in “Agent’s Characteristics” discuss whether antibiotic resistance profile is known and if so, indicate;</p> <p>10. Agents #1 and #7 - Genetically Modified - The ischemia & burn models described in Form D mention that these bacterium species express MAM. Include this information in this section and indicate whether this genetic modification was done in your lab;</p> <p>11. Agent #7 - Genetically Modified - State if production of recombinant <i>S. epidermidis</i> expressing Shinorine and Gadusol is/was performed somewhere other than your lab;</p> <p>12. Agent #8 - Genetically Modified - Modify answer to reflect the newly imported recombinant agents;</p> <p>13. Agent #11 - Since the strain is not identified by ATCC, list the ATCC accession number.</p> <p>Form C – Human and NHP Derived Materials:</p> <p>14. Section I - Established Cell Lines - Include human cell type used for lentiviral vector production;</p> <p>15. Section I - Tissues - Provide where human skin is obtained;</p> <p>16. Section II. A - Include use of human cells for production of lentiviral vector;</p> <p>17. Section II. B - Cancer models - Include whether or not human cells given to animals were previously transduced with lentiviral vector.</p> <p>Form D – Biohazard in Animals:</p> <p>18. Section I - Update your IACUC protocol number;</p> <p>19. Section II - Select "human cells" and "transduced cells" (if applicable) and complete all related fields.</p> <p>Form E – Radiation Safety Questionnaire:</p> <p>20. Update name of "X-Ray Contact Person".</p>			
	<p>Motion Approve upon modifications addressing outlined issues.</p>			
Voting Result & Dual Use	<p>YES: 11 One member did not vote</p>	<p>NO: 0</p>	<p>Abstained: 0</p>	<p>Dual Use? No</p>

IBC Protocol Number	PI	Type of Submission	Biosafety Level	IBC Items
6. 25-12-18-02	Sah	Renewal	BSL2	<i>in vivo</i> : AAV vector and Recombinant Pseudorabies Virus
IBC Requests:	<p>Main Form – General Safety:</p> <p>1. Section III. A - Centrifuge, Vortex and Pipet - Replace "hood" with "BSC";</p>			

	<div>2. Section V - Change "...they will only be...." to ...70% ethanol will only be....".</div> <div>Form D – Biohazard in Animals:</div> <div>3. Section III. C - If animal procedures are performed exclusively in LAMS, uncheck the box for "bedding dump stations" since LAMS does not have this type of equipment.</div>			
Motion	Approve upon modifications addressing outlined issues.			
Voting Result & Dual Use	YES: 12	NO: 0	Abstained: 0	Dual Use? No

V. Protocol Updates - November 26th to December 23rd (7 protocols)

- 1. IBC# 24-10-10-01 - PI: Gao - Personnel
- 2. IBC# 25-03-19-02 - PI: Plas - Personnel
- 3. IBC# 25-07-07-01 - PI: Hudock - Location
- 4. IBC# 23-10-12-01 - PI: Thompson T - Personnel
- 5. IBC# 25-08-15-01 - PI: Huaman - Personnel
- 6. IBC# 24-07-08-01 - PI: Yu - Location
- 7. IBC# 24-01-02-01- PI: Waltz - Personnel

VI. Reports

- A. IBC/BSOf (M. Espinola)
 - An NIH/OSP listening session for region 3 (Ohio) will be held on January 15th, 2026. Committee members are encouraged to register and voice their opinion.

VII. Educational Materials/Updates

- A. **Use all the tools of the trade: Building a foundation for the next era of biosecurity** Bulletin of the atomic scientists, December 2025.
- B. **Genetically Engineered Self-Spreading Vaccines for Wildlife: Promise, Case Studies, and the Precautionary Imperative**; Bulletin of the atomic scientists, December 2025.
- K. Burns adjourned the meeting at 12:16 p.m.

